

ROLE OF MEDIUM SUPPLEMENTATION OF GROWTH MODULATORS TO ALLEVIATE CADMIUM-INDUCED OXIDATIVE STRESS IN MAIZE

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Cadmium (Cd) toxicity has been a wide-spread phenomenon affecting plant metabolic functions due to excessive production of reactive oxygen species. This study was aimed at medium supplementation of optimized levels of growth modulators viz., salicylic acid (SA, 500 μ M), ascorbic acid (AsA, 500 μ M), thiourea (TU, 400 μ M) and hydrogen peroxide (H_2O_2 , 100 μ M) in improving Cd tolerance in maize (*Zea mays* L. hybrid P-1543) in sand culture under net house conditions. All treatments were applied in the growing medium by dissolving in water. Results revealed that Cd stress (1000 μ M) severely reduced the shoot and root dry mass and leaf area per plant and endogenous production of osmoprotectants (soluble sugars, free proline and glycinebetaine), phenolic compounds (soluble phenolics, flavonoids and anthocyanins) and vitamins (ascorbic acid, niacin and riboflavin) in shoot and root tissues indicating that Cd was highly toxic to the metabolic pathways. However, medium supplementation of the selected growth modulators was quite effective in reversing the Cd toxicity effect. Among the medium supplemented compounds, SA and AsA were more effective in reversing the Cd toxicity effect than TU and H_2O_2 . Presence of tight correlations of growth attributes with metabolites levels substantiated that medium supplementation was greatly effective in orchestrating the endogenous metabolite levels and supporting maize growth under Cd toxicity. A greater effectiveness of the SA, AsA and TU can be attributed to their molecular structure and physiological properties in scavenging ROS and reducing oxidative stress. The results may have great promise and practical implications for growing maize successfully in the marginally Cd contaminated soils.

Keywords: Cd toxicity, oxidative damage, ROS, stress alleviation, metabolites accumulation, phenolics.

INTRODUCTION

Oxidative stress on plants arises due to the production of reactive oxygen species (ROS), which has been identified as a major debacle in accruing the true growth and yield potential of crop plants. It has been estimated that the extent of oxidative damage ranges from as low as 7% to as high as 70% in different plant species (Wahid *et al.*, 2014). The oxidative stress results from an imbalance in production and dousing of ROS, the production being greater than dousing (Wahid *et al.*, 2014). Oxygenic photosynthesis is a major source for the production of ROS in plants (Foyer and Shigeoka, 2011).

In nature, the oxidative stress arises from sub-optimal growth conditions due to abiotic stresses such as salinity (Esfandiari and Gohari, 2017), heat (Santos *et al.*, 2017), water stress (Kar *et al.*, 2012), metal toxicity (Shahid *et al.*, 2014) etc. Likewise, disease incidence causes the hypersensitive response in plants (Etalo *et al.*, 2013). The plants deploy internal machinery to offload the excessively produced ROS, which is quite often related to induction of antioxidative enzymes, which scavenge the ROS and tend to normalize the plant metabolism (Abdelgawad *et al.*, 2017; Santos *et al.*, 2017). In addition to deployment of enzymatic antioxidative arsenal, there is a list of such non-enzymatic agents, the production of which has been quite helpful in rescuing the

cells and tissues from the ROS toxicity arising from abiotic stress factors (Hassan *et al.*, 2018; Rehman *et al.*, 2018; Sarwar *et al.*, 2018).

Nonetheless, the oxidative damage on the plants can be managed successfully with the exogenous supply of stress alleviating compounds in different modes including seed pretreatments, foliar spray and medium supplementation, although their supplementation levels may be different for different plant species (Wahid *et al.*, 2017). It is known that pre-sowing seed treatments and foliar spray have proven worthwhile in improving crop productivity by 15-20% in different crop plants by producing critical physiological and biochemical changes (Liu *et al.*, 2014; Mustafa *et al.*, 2018). The medium supplementation of different chemicals and plant extracts has been used on a limited scale but with quite promising results (Perveen *et al.*, 2015; Hussain *et al.*, 2017; Tariq *et al.*, 2018). However, there is still a need to accrue and establish the possible benefits of medium supplementation of different growth enhancing agents on firm footings especially under stressful conditions.

Among different non-essential toxic metals, cadmium (Cd) toxicity is a major one in affecting the plant growth and development throughout the world. The toxicity of Cd ranges from whole plant level to negative effects on physiological functions and disruption of the structure of macromolecules

(Perveen *et al.*, 2011; Wan and Zhang, 2012; Jibril *et al.*, 2017). The studies are imperative on the alleviation of Cd toxicity by the use of ligands and metal chelators when supplemented through growing media. Recently, Perveen *et al.* (2018) have reported the potential of medium supplemental thiourea in improving the membrane stability, osmoprotectants and vitamins synthesis and reducing the Cd toxicity on maize root and shoot tissues. Nevertheless, more studies with exogenous use of compounds of diverse nature will improve our understanding of physiological roles of these medium supplements in combating the oxidative damage and improving Cd tolerance in plants.

From the above account it is clear that Cd-induced oxidative damage is one of the major reasons of hampered plant growth and development. We assume that medium supplementation of different compounds may improve plant growth and reduce Cd toxicity by modulating the internal levels of various metabolites. In this study, an attempt was made to determine the comparative effectiveness of selected levels of medium supplemented salicylic acid (a phenolic), ascorbic acid (a vitamin), thiourea (a synthetic molecule with biological activities) and hydrogen peroxide (an oxidant) in modulating the endogenous levels of metabolites and reducing the Cd toxicity in maize seedlings.

MATERIALS AND METHODS

Plant material and experimental details: A pot experiment was done on maize (*Zea mays* L.) hybrid P-1543 (Pioneer Seeds Pakistan Ltd., Sahiwal) to investigate the comparative efficacy of selected levels of medium supplemented 0.5 mM salicylic acid (SA), 0.5 mM ascorbic acid (AsA), 0.4 mM thiourea (TU) and 0.1 mM hydrogen peroxide (H₂O₂) in improving cadmium (Cd) toxicity (1 mM). The levels of all these compounds were optimized by performing preliminary screening experiments under laboratory and greenhouse conditions in the Department of Botany, University of Agriculture, Faisalabad (data not shown). For the current experiments, 7 surface sterilized seeds were sown in pots containing 5 kg of washed river sand, and germination was completed in three days. After thinning, five uniform seedlings were kept per pot and grown for another 10 days.

Treatment application and harvesting: The above-mentioned selected levels of the selected compounds (mentioned above) were medium supplemented followed by Cd level. The potted plants were kept under bright sunlight with a ~28/~22±2°C day/night temperature, RH ~45/~58±3% and photosynthetic photo flux density of 900-980 μmol/m²/s. The design of the experiment was completely randomized with factorial arrangement having three replications (each replication constituted one pot). Five seedlings per pot were grown for seven days after abovementioned treatments. Leaf area was measured of intact plants as maximum leaf length × maximum leaf width × 0.68 (correction factor) as described

by Carleton and Foote (1965). At harvest, the plant roots were carefully removed from sand in running water. The shoots were separated from the roots, washed and blotted dried. Both the parts were put in the zip lock plastic bags and brought to the laboratory on ice. One portion of the shoot and root from three plants per replication in each treatment were put in paper bags and dried in an oven at 65°C for seven days. The other portion of the shoot and root was preserved in a freezer at -40°C until used for biochemical analysis.

Tissue Metabolites Measurement Procedures:

Oxidative stress parameters: For the estimation of H₂O₂ concentrations with the method of Velikova *et al.* (2000), the plant tissue was mixed well with trichloroacetic acid (TCA), centrifuged at 12000 × g for 15 min. A 0.5 mL of the supernatant was mixed with 0.5 mL each of 10 mM phosphate buffer and 1 M of potassium iodide; mixed well and let stand at room temperature. The absorbance of the complex was taken at 390 nm. For standard curve, a range of standards (0.2 to 1.0 mM) were prepared and run in the same manner. For the measurement of MDA with the protocol of Heath and Packer (1968), 0.1 g of fresh tissue was homogenized with 1 mL of 5% solution of TCA and centrifuged at 12000 × g for 15 min. Supernatant (1 mL) was mixed with 1 mL of thiobarbaturic acid, mixed and heated in a water bath at 95°C for 30 min, cooled and again centrifuged at 7500 × g for 5 min. The absorbance of the supernatant was taken at 532 and 600 nm. The value obtained at 600 nm was subtracted from that taken at 532 nm to correct the non-specific turbidity. To calculate MDA concentration, the extinction coefficient of 115 mM/cm was used, while 5% TCA was used as blank.

Osmoprotectants analysis: Soluble sugars were determined by reacting with the anthrone reagent (prepared by dissolving 1 g anthrone in 1 L of conc. H₂SO₄) with 1 mL of the water extracted sugars sample in a water bath at 95-100°C for 15 min. The samples were cooled in running water, vortexed and absorbance of the colored complex was taken at 620 nm (Yoshida *et al.*, 1976). The free proline was estimated using the method of Bates *et al.* (1973). A 1 mL of 3% sulphosalicylic acid extract was reacted with 1 mL acid ninhydrin (prepared by reacting 1.25 g ninhydrin with 30 mL of glacial acetic acid) and 1 mL glacial acetic acid by heating at 100°C and then the reaction mixture terminated in ice bath. The colored complex aspirated by dissolving in 4 mL of toluene. The absorbance was taken at 520 nm. A standard curve was constructed using the proline. For glycinebetaine (GB) estimation with the method of Grieve and Grattan (1983) the fresh plant material was extracted in 2 N H₂SO₄. Then 1 mL of the sample was transferred to glass tube, cooled at 4°C and reacted with cold IKI₂ (periodide) and rapidly vortexed. These tubes were kept at 4°C for 16-20 h and centrifuges at 10000 × g. The periodide crystals were dissolved in 9 mL of 1, 2-dichloroethane and vortexed till the crystals were completely dissolved and kept at room temperature for 2-2.5 h. The absorbance of the colored

solution was taken at 365 nm. A standard curve was made to determine the levels of GB in unknown samples.

Phenolic compounds estimation: The amount of total soluble phenolics was estimated with the method of Julkunen-Titto (1985). Fresh (0.1 g) plant sample was extracted with 80% acetone. An aliquot (100 µL) was diluted to 1 mL and reacted with Folin Phenol reagent by vigorous shaking. Immediately after 2.5 mL of Na₂CO₃ solution was added and made the final volume to 5 mL and mixed well. The absorbance was taken at 750 nm and tannic acid was used to prepare a standard curve for knowing the amount of soluble phenolics in unknown samples. To measure the flavonoids with the method of Zhishen *et al.* (1999), 0.1 g of the plant material was extracted in 80% acetone. Then to 1 mL of extract was added to 4 mL of water in a 10 mL volumetric flask. After 5 min., 0.6 mL of 5% NaNO₂ followed by 0.5 mL of 10% AlCl₃ solution were added and mixed. After 1 min, 2 mL of 1 M NaOH was added and made the volume up to 10 mL. The absorbance was taken at 520 nm while using 80% acetone as blank. For the measurement of anthocyanins with the method of Stark and Wray (1989), 0.1 g of fresh material was extracted with 1% acidified methanol and heated in a water bath at 50°C and filtered the extract. The absorbance of the supernatant was taken at 535 nm while methanol was used as a blank.

Vitamins analysis: for the determination of AsA with the method of Mukherjee and Choudhuri (1983), 0.1 g of the plant material was homogenized with 2 mL of 6% TCA and filtered. A 1 mL of the extract was reacted with 1 mL of the dinitrophenyl hydrazine followed by a drop of 10% thiourea solution (prepared in 70% ethanol). After heating the mixture was cooled in an ice bath for 15 min. While still on ice 5 mL of 80% H₂SO₄ was added; mixed and absorbance was taken

at 530 nm, A graded series of AsA solution was prepared to make standard curve. For the estimation of niacin with the method of Okwu and Josiah (2006), 0.1 g of fresh plant material was added to 1 mL of 1 N H₂SO₄ and shaken well for 20 min and then added 1 drop of ammonia solution and filtered. Then 1 mL of the filtrate was reacted with 0.5 mL of 10% KCN and 0.5 mL of 0.02 N H₂SO₄ and shaken. The absorbance of the yellow colored complex was taken at 470 nm. Niacin (Sigma) was used to construct a standard curve. Riboflavin level was measured as described by Okwu and Josiah (2006). A 0.5 g of fresh plant sample was extracted with 10 mL of 50% ethanol and filtered. To 1 mL of extract, 1 mL each of 5% solution of KMNO₄ and 30% H₂O₂ were added and heated in a water bath at 50°C for 30 min. When cool, 0.2 mL of 40% Na₂SO₄ was added and final volume made up to 5 mL and mixed well. The absorbance of the reaction mixture was taken at 510 nm. Riboflavin (Sigma) was used to prepare a standard curve.

Statistical analysis: Statistix8.0 program was used to carry out analysis of variance of the data recorded for shoot and root dry mass and leaf area per plant, and metabolites to find out significant differences among the stresses and medium supplementation treatments. Least significant difference (LSD) test was used to find out delineate the differences among the treatments (Steel *et al.*, 1996).

RESULTS

Growth parameters: Statistical analysis of data indicated significant differences in the stress treatments and medium supplemented compounds both in the dry mass of shoot and root and leaf area per plant (Table 1). Compared with control, the medium supplementation of SA was the most effective in

Table 1. Critical value for comparison and mean squares of two way interactions of growth and physiological attributes of maize with the medium supplementation of stress mitigating agents under control and Cd toxicity conditions.

Parameters	Shoot		Root	
	CV for comparison	Stress × treatment interaction	CV for comparison	Stress × treatment interaction
Dry weight	0.021	0.16×10 ⁻³ **	0.014	0.07×10 ⁻³ **
Leaf area per plant	0.963	0.320*	-	-
Hydrogen peroxide	1.144	0.451**	1.509	0.785**
Malondialdehyde	1.198	0.494**	1.018	0.357**
Soluble sugars	1.408	0.684*	1.244	0.533*
Free proline	2.095	1.513**	3.050	3.206**
Glycinebetaine	1.650	0.938**	0.880	0.267**
Soluble phenolics	5.226	9.410*	5.045	8.770*
Flavonoids	1.560	0.860**	0.942	0.850**
Anthocyanins	0.048	0.81×10 ⁻³ **	0.045	0.70×10 ⁻³ **
Ascorbic acid	1.206	0.501**	0.737	0.187**
Niacin	0.759	0.197**	0.456	0.072**
Riboflavin	0.389	0.052**	0.596	0.123**

Significant at *, P<0.05 and ** P<0.01

enhancing shoot dry weight both under control condition followed by thiourea and AsA while H₂O₂ was ineffective in this regards. However, under Cd stress, the shoot dry weight was highly reduced but all the medium supplements enhanced this attribute although SA was the most effective followed by AsA, TU and H₂O₂ (Fig. 1a).

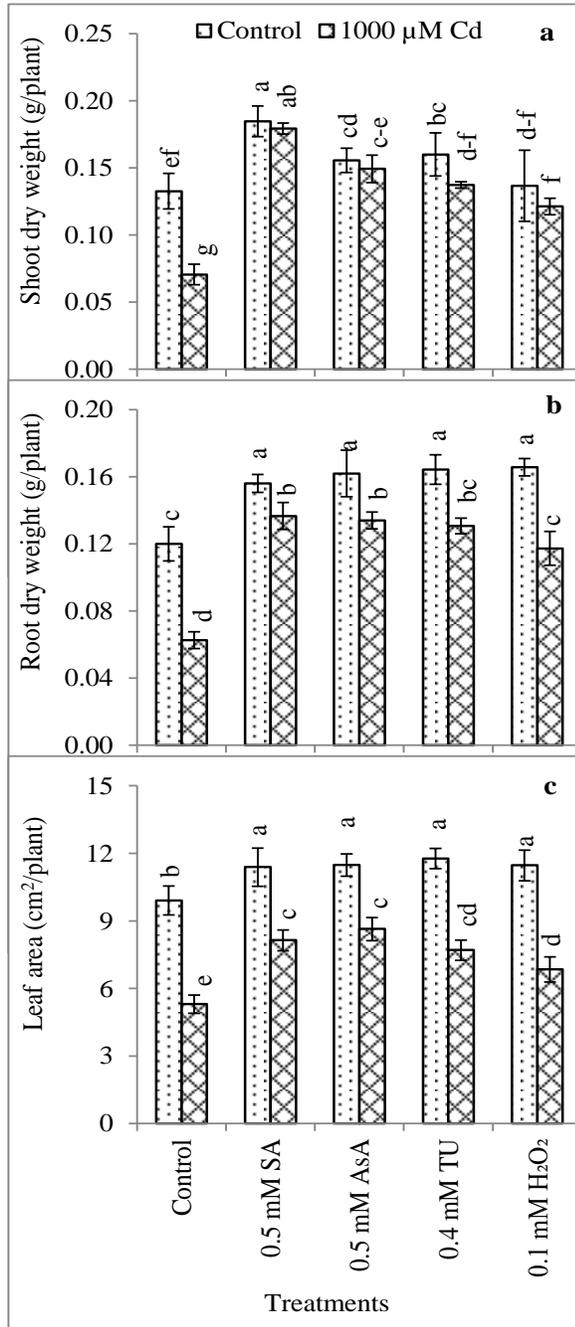


Figure 1. Growth attributes of maize seedlings with medium supplementation of selected levels of stress alleviating compounds under Cd stress.

The root dry weight, being low under control condition, was equally improved with the medium supplementation of all the chemicals. However, under Cd stress, the improvement in this attribute was the highest with SA and AsA followed by TU and H₂O₂ (Fig. 1b). As regards leaf area per plant, the medium supplemented with SA, AsA and TU indicated a similar improvement while H₂O₂ was not much effective. However, under Cd intoxication, although leaf area per plant was not improved much yet TU was more effective followed by SA and AsA while H₂O₂ was not effective (Fig. 1c).

Changes in Tissue Metabolite Levels:

Oxidative stress parameters: Data showed significant differences in the stress and medium supplementation treatments with significant interaction of these factors for shoot and root H₂O₂ and MDA (Table 1). The shoot H₂O₂ contents were the lowest under control condition, which indicated some increase with the medium supplementation of all the compounds. However, Cd stress produced a twofold increase in shoot H₂O₂ contents while medium supplementation of all the selected compounds substantially reduced the H₂O₂ content; TU being relatively more effective followed by H₂O₂ (Fig. 2a). As regards root, the H₂O₂ level was the lowest in under medium supply of SA but relatively higher under H₂O₂ supplementation. However, Cd treatment caused a 2.2-folds increase in H₂O₂ in root while all the medium supplemented compounds were almost equally effective in reducing the production of H₂O₂, although SA and AsA were relatively more effective (Fig. 2b). As regards MDA contents, under no Cd stress the medium supplementation of SA was the most effective followed by H₂O₂ in reducing the shoot MDA contents. However, Cd treatment of maize led to about 2.5 folds greater increase in shoot MDA while among the medium supplemented compound SA was the most effective followed by AsA (Fig. 2c). As for root tissue the MDA content was the lowest under non-medium supplemented plants, which were little higher with all the medium supplements. However, with Cd treatment, the non-supplemented plants indicated the highest MDA content, which declined substantially with all the medium supply, although SA was the most effective followed by TU (Fig. 2d).

Osmoprotectants accumulation: Statistical analysis of data for tissue contents of soluble sugars, free proline and GB indicated significant differences of Cd stress and medium supplementation along significant interaction of these factors (Table 1). As regards soluble sugars, under control condition, the medium supply of SA was the most effective in enhancing in the shoot soluble sugar contents while TU and H₂O₂ were not effective at all. However, under Cd stress there was a reduction in the soluble sugar content while medium supplementation of SA and AsA at selected levels were quite effective in enhancing the soluble sugar contents (Fig. 3a).

The root tissue exhibited lower soluble sugars contents under which was improved by 2.5 times with medium supply of SA

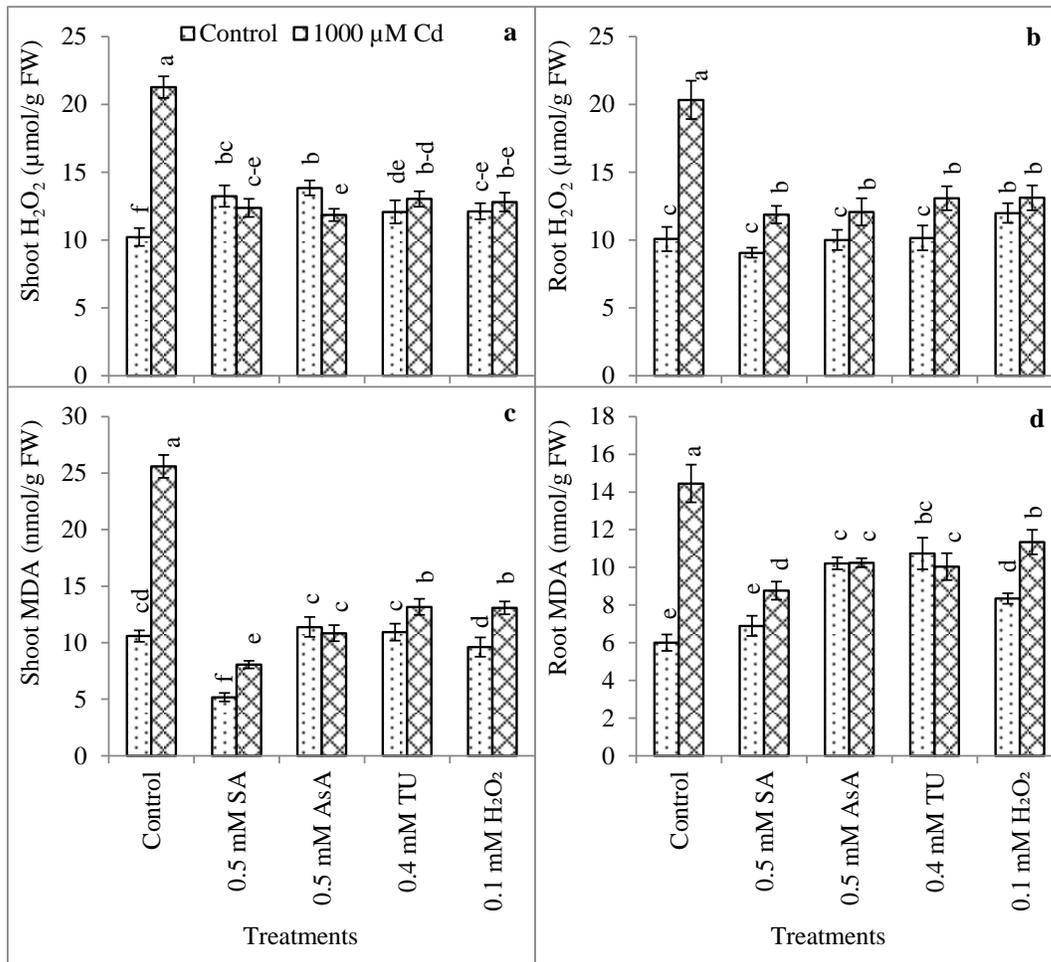


Figure 2. Changes in the tissue levels of hydrogen peroxide (H₂O₂) and malondaldehyde (MDA) in Cd-intoxicated maize seedlings with the medium supplementation of selected levels of stress alleviating compounds.

control while medium supplementation of AsA was the most effective in improving this attribute. Contrarily under Cd stress, medium supply of TU and SA were the most effective in improving the root soluble sugar contents (Fig. 3b). As regards free proline accumulation, shoot free proline was lower under no medium supplementation treatments while under Cd stress both SA and AsA manifested about twofold greater accumulation of free proline followed by H₂O₂ and TU (Fig. 3c). Root free proline accumulation did not differ much with or without medium supplementation treatments. However, under Cd stress the root free proline content displayed a twofold increase with SA and AsA while 1.5 folds increase with TU and H₂O₂ when compared with no-medium supplementation (Fig. 3d). Under no-Cd treatment, medium supplementation with AsA was the most effective in improving shoot GB followed H₂O₂ compared with other compounds. However, Cd intoxication reduced the shoot GB,

and by two times with AsA, TU and H₂O₂ (Fig. 3e). As regards root GB, under no Cd toxicity but with medium supplementation the increase in root GB was in the order SA = AsA > H₂O₂ > TU. On the other hand, the Cd induced reduction in root GB was improved by about two times with SA and AsA but by ~1.5 times with H₂O₂ and TU (Fig. 3f).

Phenolics accumulation: Data indicated significant difference in and the interactions of the medium supplemented compounds for soluble phenolics, flavonoids and anthocyanons accumulation in shoot as well as root tissue (Table 1). Except for SA under control condition, which showed greater shoot soluble phenolics, all other supplemental compounds did not differ from each other. However, under Cd stress there was more than two times higher accumulation of shoot soluble phenolics with SA and AsA followed by TU and H₂O₂ (Fig. 4a).

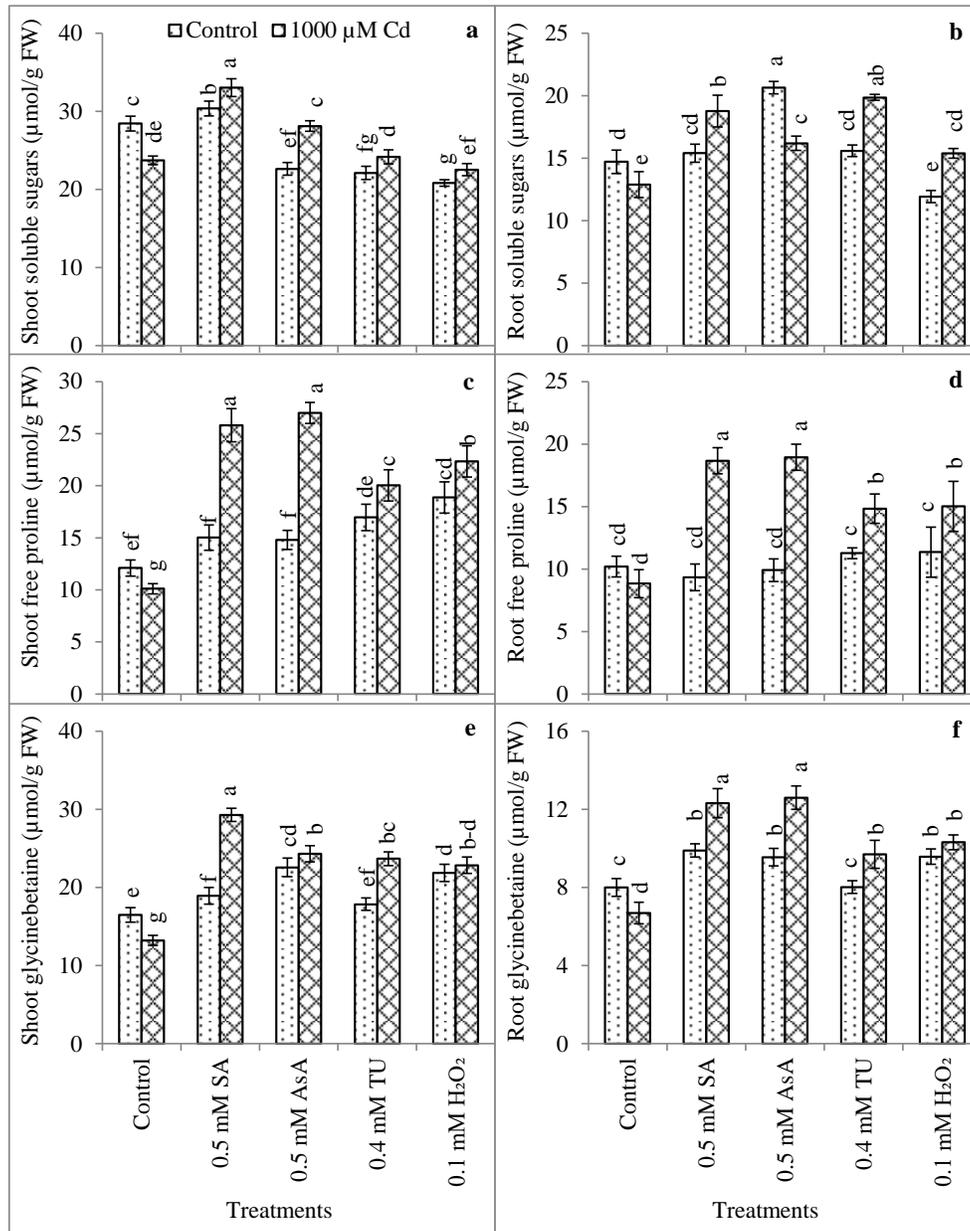


Figure 3. Changes in the tissue levels of hydrogen peroxide (H₂O₂) and malondaldehyde (MDA) in Cd-stressed maize seedlings with the medium supplementation of selected levels of stress alleviating compounds.

As for root soluble phenolics, the data revealed that under no Cd stress, except for an increase in soluble phenolics under SA supplementation, there was no difference in control and rest of the medium supplementations. Conversely under Cd toxicity, medium supplementation with SA and AsA led to a greater soluble phenolics accumulation followed by TU and H₂O₂ (Fig. 4b). Under no Cd toxicity the shoot flavonoids kept low irrespective of the medium supplementation treatments. However, under Cd stress, the shoot flavonoids

were the highest with medium supply of SA and AsA but low with TU and H₂O₂ (Fig. 4c). The root flavonoid contents remained low with or without medium supplementations under no Cd stress. However, under Cd toxicity, SA followed by AsA exhibited the greatest increase in the root flavonoids followed by TU and H₂O₂ (Fig. 4d). Under no Cd treatment the shoot anthocyanins contents were the highest with medium supplementation of SA followed by AsA. On the other hand under Cd stress, highest shoot anthocyanins were

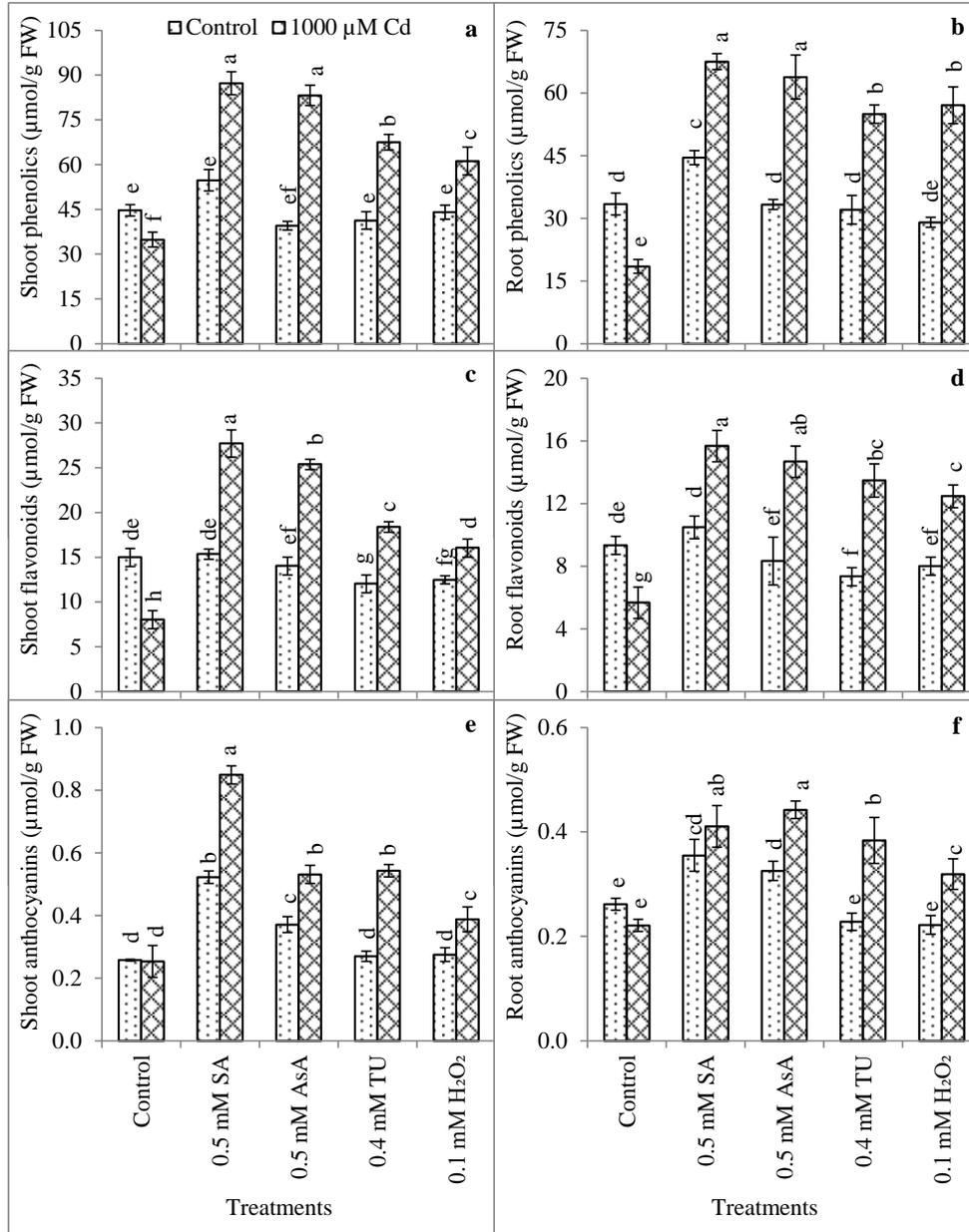


Figure 4. Variations in the tissue levels of soluble phenolics, flavonoids and anthocyanins contents in Cd-treated maize seedlings with the medium supplementation of selected levels of stress lessening compounds.

analyzed in SA supplementation (three times higher than the Cd treated controls) while in the rest of the treatments the anthocyanins contents were quite low in concentration (Fig. 4e). In root tissue under no Cd stress, the anthocyanins contents were significantly higher with medium supply of SA followed by AsA. However, under Cd stress, medium supplementation caused generally greater shoot anthocyanins being the highest with SA and AsA supplementation followed by TU and H₂O₂ (Fig. 4f).

Vitamins biosynthesis: Results revealed significant differences in the levels of AsA, niacin and riboflavin under Cd stress and medium supplementation of various compounds (Table 1). Under no Cd toxicity the shoot AsA level was the highest in the plants supplemented with AsA while its level was almost similar to control in rest of the medium supplementation treatments. Under Cd stress, however, the shoot AsA was the lowest in the non-medium supplemented plants while the highest with the medium supplementation of SA and AsA (Fig. 5a).

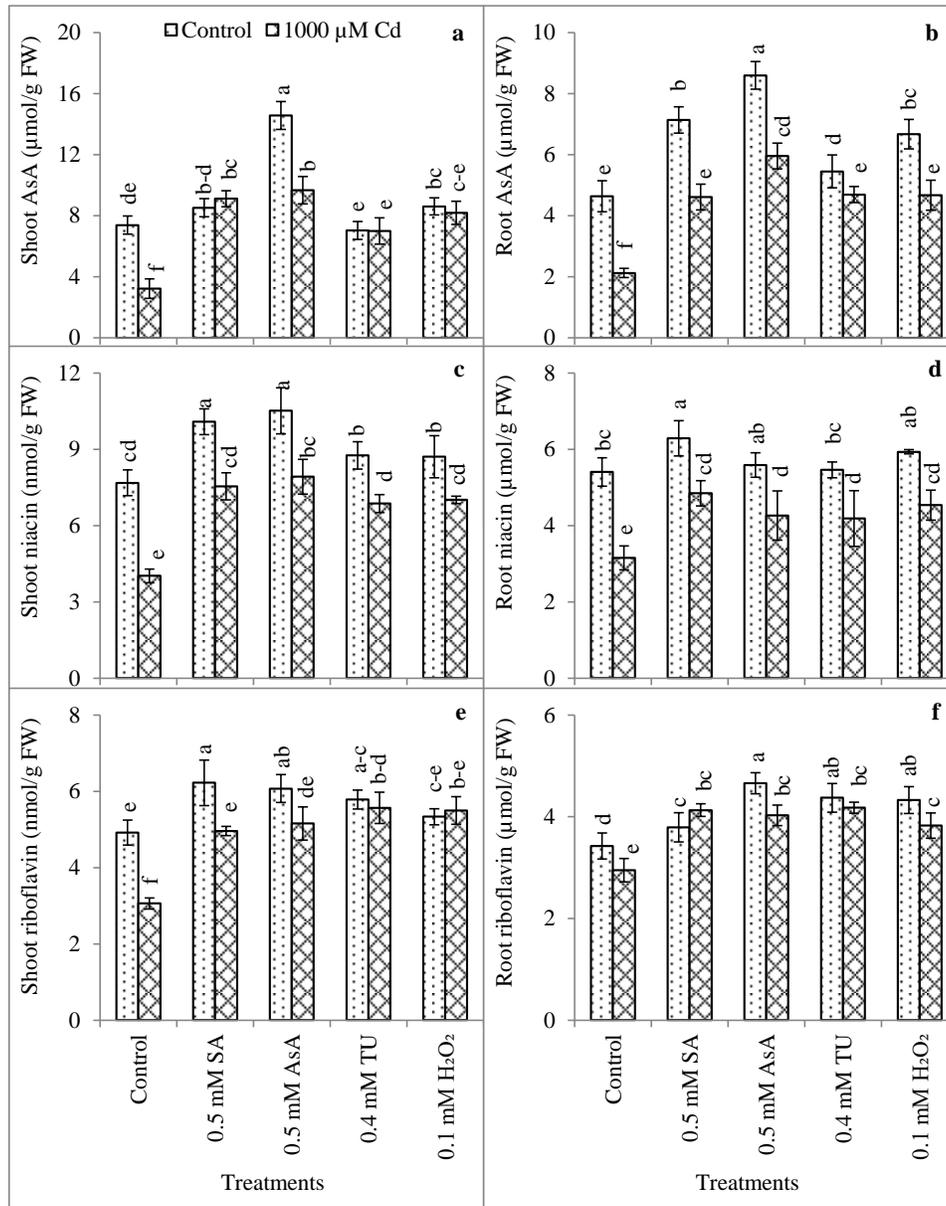


Figure 5 Variations in the tissue levels of ascorbic acid, niacin and riboflavin in the shoot and root of Cd-treated maize seedlings with the medium supplementation of selected levels of stress mitigating compounds.

In root tissue the medium supplementation with AsA led to a highest accumulation of AsA followed SA and H₂O₂. The trend of AsA accumulation was higher noticed under Cd stress (Fig. 5b). As regards niacin contents, under control (no Cd stress), the shoot niacin was the highest with medium supplementation of AsA and SA followed by TU and H₂O₂. However, under Cd stress, shoot niacin, being lowest in untreated plants, accumulated it highly in SA and AsA treated plants (Fig. 5c). As regards root niacin contents, under no-Cd treatment, the medium supplementation of compounds did not

show difference from the control plants. However, under Cd stress, medium supplementation with SA and H₂O₂ were comparatively more effective as compared to other treatments (Fig. 5d). Under control (no Cd) condition the shoot riboflavin contents were almost similar with the medium supplementation of SA, AsA and TU. However, under Cd stress the shoot riboflavin was the lowest in non-medium supplemented plants but was the highest in TU supplemented plants followed by H₂O₂ and AsA supplements (Fig. 5e). As for root riboflavin contents, under control (no Cd stress)

Table 2. Pearson's correlation coefficient (*r*) of some growth attributes with some physiological attributes of maize hybrids under medium supplementation of thiourea subjected to control and Cd stress conditions.

Variable	Shoot dry weight		Leaf area per plant		Root dry weight	
	Control	Cd stress	Control	Cd stress	Control	Cd stress
Hydrogen peroxide	0.587ns	-0.879*	0.736ns	-0.903*	0.254ns	-0.988**
Malondialdehyde	-0.748ns	-0.964**	-0.114ns	-0.917*	0.744ns	-0.969**
Soluble sugars	0.442ns	0.779ns	-0.546ns	0.626ns	0.113ns	0.822ns
Free proline	0.056ns	0.926*	0.777ns	0.933*	0.313ns	0.938*
Glycinebetaine	-0.083ns	0.977**	0.564ns	0.879*	0.521ns	0.906*
Soluble phenolics	0.657ns	0.990**	-0.095ns	0.969**	-0.402ns	0.979**
Flavonoids	0.272ns	0.984**	-0.578ns	0.946**	-0.459ns	0.992**
Anthocyanins	0.868ns	0.944*	0.280ns	0.769ns	-0.017ns	0.946*
Ascorbic acid	-0.037ns	0.905*	0.259ns	0.915*	0.618ns	0.918*
Niacin	0.616ns	0.915*	0.627ns	0.944*	0.370ns	0.895*
Riboflavin	0.659ns	0.901*	0.649ns	0.895*	0.849ns	0.989**

Significant at *, $P < 0.05$; **, $P < 0.01$ and ns, $P > 0.05$

condition, the value of this attribute was the highest with medium supplementation of AsA, TU and H₂O₂. However, under Cd stress maximum root riboflavin was analyzed with medium supply of SA, AsA and TU (Fig. 5f).

Correlations: To validate the possible association of different metabolite levels with the growth attributes (shoot and root dry mass and leaf area per plant), the Pearson's correlation coefficients were computed. Under control conditions shoot dry mass, leaf area per plant and root dry weight were not correlated with any of the metabolites. However, under Cd stress, all the growth parameters were significantly correlated with the metabolites except no correlation of soluble sugars with any of the growth attributes and no correlation of leaf area with anthocyanins contents (Table 2).

DISCUSSION

In this study, it was noted that Cd stress adversely affected the growth of maize, while the medium supplementation of the selected levels of compounds minimized the adverse effect of Cd, although their effectiveness was variable (Fig. 1). It has been reported that Cd stress applied to plant roots competes with the calcium uptake at Ca channels, competitively taken up and causes toxicity to the cytoplasmic structures (Wahid *et al.*, 2009; Choppala *et al.*, 2013). In this research we noticed that there was a reduction in the dry weight of shoot and root as well as leaf area per plant, while the medium supplementation of SA, AsA, TU and H₂O₂ helped the plants to diminish the toxic effect of Cd as noticed from markedly greater dry weight of shoot and root and leaf area per plant of maize (Fig. 1). The major effect of Cd as toxic metal is the production of ROS, leading to the disruption of lipid bilayer in the membranes. This can be measured in terms of MDA contents as a consequence of peroxidation of membrane lipids (Shahid *et al.*, 2014; Kumar *et al.*, 2018). So, a linear relationship exists between the ROS and MDA content (Tian

et al., 2012). Our data suggested that due to Cd toxicity, both H₂O₂ and MDA were produced in quite high concentrations (Fig. 2), but the medium supplementation of compounds used in this study have definitive roles in rescuing the plants from Cd damage.

Various metabolites produced endogenously protect the membranes and improve stress tolerance. Major ones include osmoprotectants, phenolics and vitamins. All these compounds have their specific modes of action (Emamverdian *et al.*, 2015; Belkadhi *et al.*, 2016; Tariq *et al.*, 2018). In view of the fact that membrane damage is a major effect of heavy metal toxicity, it was deemed appropriate to find out the possible mechanistic basis of medium supplemented compounds in lessening the adverse effect of Cd on the shoot and root tissues. The matter of specific interest for us was to find out changes in all these compounds with the medium supplementations. Our results suggested that with Cd intoxication, there was an enormous reduction in the levels of soluble sugars, free proline and GB (Fig. 3), decline in the levels of soluble phenolics and anthocyanins among the flavonoids (Fig. 4) and also reduced the production of AsA, niacin and riboflavin among the vitamins (Fig. 5). This indicated that Cd was entirely toxic to metabolic pathways probably inhibiting the enzymatic activities (Hassan *et al.*, 2013). However, medium supplementation of the selected compound improved their tissues concentrations to a great extent. Among the medium supplemented compounds, SA was most effective followed by AsA and TU while H₂O₂ was the least effective (Fig. 3-5). It was quite evident from the results that improved growth of maize with the medium supplementation was due to concerted action of the endogenously synthesized metabolites, which included protection of cellular membranes (by osmoprotectants), scavenging of ROS (phenolic compounds) and sustained metabolic activities (vitamins). So, these results indicated that Cd toxicity was effectively minimized by the exogenous application of different growth enhancing compounds.

To further substantiate the above findings the correlations were drawn between the changes in growth attributes and metabolite levels under control and Cd toxicity conditions (Steel *et al.*, 1996). The data revealed that under control conditions none of the growth attributes indicated any relationship with any of metabolic parameters. However, under Cd toxicity MDA and H₂O₂ were negatively while the rest of the attributes including osmoprotectants, phenolics and vitamins were positively related to shoot and root dry weight and leaf area per plant. These data confirmed that the medium supplementation have effective role not only in mitigating the adverse effect of Cd but also normalizing the metabolic functions in maize. Among the medium supplemented compounds, SA followed by AsA were the most effective. Thus, the medium supplementation of the chemical used in the present research emerged as an effective tool in growing maize at least in marginally cadmium contamination. Greater effectiveness of the SA, AsA and TU are assignable to their molecular and physiological properties in scavenging ROS.

Conclusion: The Cd-intoxication of maize seedlings caused oxidative damage on the plant growth attributes especially disrupting the metabolic functions as revealed from much reduced levels of osmoprotectants, phenolics and vitamins while medium supplementations competently improved the endogenous levels of all these metabolites. Among the medium supplemented compounds, SA and AsA were more effective by virtue of their prominent metabolic role as effective ROS scavengers in plants. The results carry great implications for successfully growing maize in marginally Cd-contaminated soils.

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