

PRODUCTION OF SINGLE CELL PROTEIN FROM DELIGNIFIED CORN COB BY *Arachnietus* species

M. Javed Asad, M. Asghar, M. Yaqub & Khurram Shahzad
Department of Chemistry, University of Agriculture, Faisalabad

The study was carried out to utilize alkali treated corn cob as a substrate for single cell protein production by *Arachnietus* sp. The maximum crude protein (18.87 %) was obtained after 96 hr of continuous shaking (120 rpm) fermentation of 2% NaOH treated corn cob by *Arachnietus* sp. The optimum culture medium of alkali treated corn cob (3%) contained (g/100 ml): urea, 2; CaCl₂ · 2H₂O, 0.05; MgSO₄ · 7H₂O, 0.10; and KH₂PO₄, 0.3. Crude protein content of alkali treated corn cob (inherently containing 3.2% crude protein) was found to increase up to 11.1% within four days. This biomass product may be used as a protein supplement in the poultry and livestock rations which presently are very costly due to the use of conventional ingredients such as oilseed cakes.

Key words: *Arachnietus* sp., corn cob, single cell protein

INTRODUCTION

A huge population is suffering from protein malnutrition in the third world and most probably by the end of second decade of this century, the food requirements would be doubled. Animal feed industry is also passing through an era of inadequate and high cost availability of conventional ingredients such as oilseed cakes and certain cereals. Also, there is a strong competition between human beings and livestock for conventional protein sources. Supplementation of vegetable proteins with animal proteins results in higher feeding costs (Dasilva et al., 1987). It is therefore imperative to produce economical quality proteins from non-conventional sources.

Microorganisms are capable of utilizing the organic matter of such residues as a source of energy for their growth. They require carbon compounds for the synthesis of cell biomass for which they have the ability to convert inorganic nitrogen into their body proteins (Khan, 1992). The protein from microorganisms is cheap and competitive with other protein sources. It may have good nutritive value depending, however, upon amino acid composition.

Such low cost agro-industrial lignocellulosic wastes must be treated by physical/chemical methods to liberate cellulose from lignins, since cellulose in lignin-hemicellulose-cellulose complex is not accessible to enzymatic hydrolysis (Bungay, 1982). Corn cob is a very hard lignocellulosic waste of corn industry and cannot be utilized efficiently by microbes without pretreatment. The present study was conducted to delignify corn cob by NaOH pretreatment and utilize it as a substrate for single cell protein (SCP) production by *Arachnietus* sp.

MATERIALS AND METHODS

Substrate: An agro-industrial waste, corn cob, procured from the CPC Rafhan, Faisalabad, was dried in an oven at 100°C to a constant weight. The dried substrate was

ground (40 mm mesh), soaked in 2% NaOH for 24 hr in 1:20 ratio and used in fermentation medium without washing.

Organism: *Arachnietus* sp. obtained from the Department of Plant Pathology, was raised on substrate-agar slants sporulation medium (Bajwa et al., 1991) and incubated at pH 4 and 32°C for 72 hr for sporulation.

Inoculum: The inoculum medium was prepared (Bajwa et al., 1991) and its pH was adjusted at 4 with MCH₃COOH/M NaOH. It was sterilized by autoclaving at 121°C (1.1 kg/cm²) for 15 minutes. Spores of the fungus were transferred into it from sporulation medium and the flask was incubated for 72 hr on shaker (120 rpm).

Aerobic Fermentation: Duplicate Erlenmeyer flasks (500 ml) containing 100 ml growth medium (to be tested) were prepared and their pH was adjusted at 4 (with MCH₃COOH/M NaOH). The flasks were plugged with cotton and autoclaved for 15 minutes. Spore inocula (51111) containing 10⁸ spores/ml, were added aseptically to each flask in laminar air flow with the help of a sterilized pipette. The flasks were then incubated at 32°C on rotary shaker (120 rpm) for aerobic fermentation. The studies were conducted systematically in such a way that the condition optimized in one experiment was maintained in the subsequent investigations.

Harvesting: Fermented samples were harvested by steaming for 5 minutes and filtered. The residues (biomass) were dried in an oven at 100°C to a constant weight, homogenized and analyzed for crude protein content.

Analytical Method: Nitrogen content of the biomass was estimated colorimetrically (Oser, 1976), using Nessler's reagent on spectronic 21 at 400 nm by comparing the absorbance of the samples with that of standard (NH₄)₂SO₄ solution. The crude protein content was calculated by the following formula:

Crude protein (%) = N% × 6.25

RESULTS AND DISCUSSION

Optimization of Fermentation Parameters

Fermentation Period: The results revealed that biomass produced after 24, 48, 72, 96 and 120 hr of incubation contained 8.02, 10.15, 10.67, 11.20 and 10.75% crude protein respectively (Table I). It was observed that maximum biomass protein (11.20%) was produced after 96 hr of incubation and gradually decreased thereafter, up to 120 hr. Similar results have been reported by Bajwa et al. (1991) who produced maximum crude protein from alkali treated rice straw by *Archniotus* sp. with shaking at 30°C for 48 hr under optimum conditions.

Table 1. Crude protein content of biomass produced by *Archniotus* sp. at different fermentation periods and varying concentrations of substrate and urea

Sr. No.	Fermentation period (hr)	Substrate level (%)	Urea	Crude protein (%)
1	24			8.02
	48			10.15
	72			10.67
	96			11.20
	120			10.75
2		1		9.47
		2		11.22
		3		13.39
		4		12.80
		5		12.16
3			0.0	13.24
			1	14.36
			2	15.48
			3	14.87
			4	13.65
			2	

Table 2. Crude protein content of biomass produced by *Archniotus* sp. with varying concentrations of KH_2PO_4 under optimum conditions

KH_2PO_4 (%)	Crude protein (%)
0.0	17.42
0.1	17.64
0.2	17.95
0.3	18.87
0.4	18.37

* Substrate 3%, urea, 2%, $CaCl_2 \cdot 2H_2O$ 0.05%; $MgSO_4 \cdot 7H_2O$ 0.01%; pH 4 and 32°C

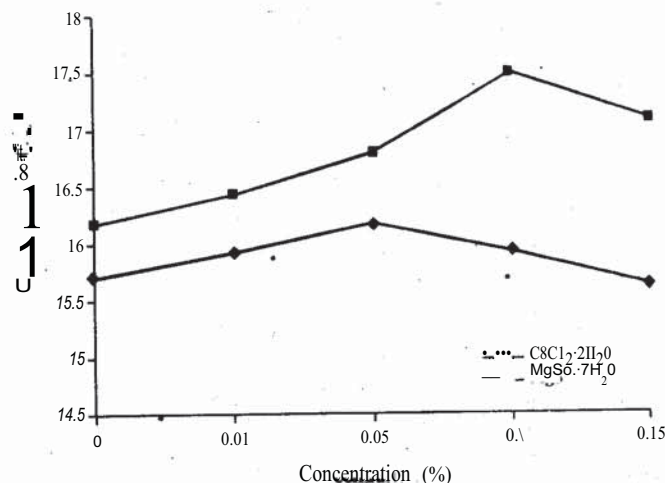


Fig. 1. Effect of varying concentration of $CaCl_2 \cdot 2H_2O$ and $MgSO_4 \cdot 7H_2O$ on SCP production.

Substrate Level: Results regarding the crude protein content of the biomass produced with different substrate levels have been given in Table I. It was noted that with an increase in substrate level the crude protein content of the biomass increased and reached a maximum of 13.19% with 3% substrate. These results are supported by Alam (1986) who reported maximum crude protein (17.2%) production by *Archniotus* sp. after 72 hr incubation with 5% (w/v) rice polishing.

Urea: It was observed that addition of urea enhanced SCP production by *Archniotus* sp. Addition of 1, 2, 1 and 4% urea to the fermentation medium under optimum conditions resulted in 13.96, 15.48, 14.87 and 11.65% crude protein respectively after 96 hr. The results are in agreement with those of Moo-Young et al. (1992) who used different nitrogen containing compounds such as (NH_3) , SO_4 , 0.47g and urea 0.86g and observed maximum crude protein (31.1%) in the medium containing urea as a nitrogen source.

$CaCl_2 \cdot 2H_2O$: The results in terms of crude protein percentage have been presented in Table 1. It was observed that addition of $CaCl_2 \cdot 2H_2O$ had favourable effect on SCP production. Crude protein content of biomass increased from 15.39 to 16.24% and 0.05% $CaCl_2 \cdot 2H_2O$ produced higher biomass protein than control (without $CaCl_2 \cdot 2H_2O$) and all other concentrations of this salt. These results are in accordance with those of Chahal et al. (1987) who produced maximum biomass protein (40%) from corn stover by *Pleurotus sajorajju* with 0.03% $CaCl_2 \cdot 2H_2O$ along with optimum concentrations of other micronutrients.

$MgSO_4 \cdot 7H_2O$: The results indicated that 0.1% $MgSO_4 \cdot 7H_2O$ in the medium gave the highest SCP yield. The crude protein content of the biomass increased with the addition of $MgSO_4 \cdot 7H_2O$ and reached its maximum (17.42%) at 0.1% level under optimum conditions.

Production of single cell protein

Hongpattarakere and Kittikun (1995) fermented cassava by *Schwanniomyces castellii* and reported 0.1% MgSO₄·7H₂O as optimum for the production of maximum biomass protein (7.4 g/100 g of starch).

KH₂PO₄; The maximum crude protein yield (18.87%) was recorded with 0.3% KH₂PO₄ in the medium which was considered as the optimum level of this salt. The results of this study are in line with those of Hashmi et al. (1991) who also reported 0.2% KH₂PO₄ as optimum for maximum SCP production by *Arachniotus* sp. Aalia et al. (1998) reported an increase in crude protein with addition of 0.3% KH₂PO₄ to corn stover medium fermented with *Arachniotus* sp.

REFERENCES

- Aaha, S., M. Rashid, M. Asghar and M. Yaqub. 1998. Bioconversion of corn stover into biomass protein by *Arachniotus* sp. Pak. J. Biol. Sci. 1(2):78-80.
- Alam, R.S. 1986. Production of basic amino acids by *Arachniotus* sp. M.Sc. Thesis. Univ. Agri. Faisalabad.
- Bajwa, M.A., T. Aziz and A.S. Hashmi. 1991. Production of fungal biomass protein from alkali treated rice straw by *Arachniotus* sp. JAPS. 1(2):79-81.
- Bungay, H.R. 1982. Biomass refining. Science. 218:643-646.
- Chahal, D.S., M. Ishaque, D. Broiullard, E. Chornct, R.P. Overend, L. Jaulin and I. Bouchard. 1987. Bioconversion of hemicelluloses into fungal protein. J. Microbiol. 78:355-361.
- Dasilva, E.L., Y.R. Dommergues, E.J. Nyns and C. Ratbidge. 1987. Microbial Technology in the Developing World. Oxford Univ. Press. Oxford. U.K.:239.
- Hashmi, A.S., K.K. Batajoo and M.A. Bajwa. 1991. Bioconversion of rice straw into protein concentrate with *Arachniotus* sp. Proc. Int. Symp. Biotechnology for Energy 16-21 Dec. 1989. Faisalabad, Pakistan: 149-155.
- Hongpattarakere, T. and A.H. Kittikun. 1995. Optimization of single cell protein production from cassava starch using *Schwanniomyces castellii*. World J. Microbiol. Biotechnol. 11:607-609.
- Khan, M.Y., M.U. Dahat and M.Y. Khan. 1992. Enzyme and fermentation biotechnology. Proc. All Pak. Sci. Conf. 16-21 May. Khanspur, Pakistan.
- Moo-Young, M.Y., Chish and D. Vlach. 1992. Fermentative conversion of cellulosic substrates into microbial protein by *Neurospora sitophila*. Biotech. Letter. 14(9):X63-K6K.
- Oser, B.L., 1976. Hawk's Physiological Chemistry. Tata McGraw Hill Publishing Co., New Delhi: 1218.